

Denitrification in a Chinampa soil of Mexico City as affected by methylparathion

Blanco-Jarvio A.^a, Chávez-López C.^a, Luna-Guido M.^b, Dendooven L.^b, Cabirol N^{c,*}

^a Instituto de Ingeniería, Coordinación de Bioprocesos ambientales, Universidad Nacional Autónoma de México (UNAM), Mexico city,

^b Laboratorio de Ecología de Suelos, GIB, Cinvestav, Mexico City,

^c Facultad de Ciencias, Departamento de Biología Celular, Universidad Nacional Autónoma de México (UNAM), Mexico City.

Abstract

Chinampas are raised garden beds used for agriculture in the southeastern part of the Valley of Mexico since pre-Hispanic times. In modern times, large amounts of pesticides, such as methylparathion, have been used with an unknown effect on soil processes, such as denitrification and the ratio of N₂O-to-N₂. Soil from a Chinampa was amended with nitrate (NO₃⁻) with or without chloramphenicol known to inhibit *de novo* synthesis of enzymes, with or without acetylene (C₂H₂) known to inhibit the reduction of nitrous oxide (N₂O) to dinitrogen (N₂) and spiked with or without methylparathion. Methylparathion increased the concentration of NO₂⁻ and removal of NO₃⁻ from soil, but increased the emission of N₂O and N₂. The concentration of NO₂⁻, the removal of NO₃⁻ production, and the N₂O and N₂ emission rates were generally larger when the aerobic conditioning of the soil was less than 14 days. It was found that methylparathion increased the denitrification process and showed no inhibitory effects on emissions of N₂O or N₂.

Key words

Chloramphenicol; dynamics of mineral N; nitrous oxide to dinitrogen ratio; organophosphorous pesticide

Introduction

In Xochimilco (Mexico City), agriculture is done in a unique way called '*Chinampa*' since pre-Hispanic times. *Chinampa* is a pre-Columbian form of agriculture whereby sediment from canals is collected regularly and applied to human-made islands forming raised garden beds called chinampas. They contributed greatly to the development of indigenous cultures that occupied the southeastern part of the Valley of Mexico, i.e. Tenochtitlan. The soils are deep and discontinuous and due to the human influence some authors classify them as Anthrosols (INECOL 2002).

Chinampas are intensively cultivated whereby different pesticides, such as clorpiriphos, methylparathion, malathion, lindane and atrazine, are used in large quantities (CICOPLAFEST 2008). Little information is available how methylparathion might affect soil processes. However, Zhang *et al.* (2006) found that in a soil contaminated with methylparathion the diversity and structure of microbial communities changed.

Negative effects on non-targeted soil microorganisms have been reported when pesticides are applied and microbial activity and diversity is reduced in soil (Johnsen *et al.* 2001; Locke and Zablutowicz 2004; IPCC 2007b). Nevertheless, other studies found that fumigation increased organic matter degradation rates (Locke and Zablutowicz 2004). Recently, it has been reported that fumigation with pesticides increased nitrous oxide (N₂O) emissions (Spokas and Wang 2003; Spokas *et al.* 2005; Spokas *et al.* 2006). This suggests that pesticides might affect the denitrification process. However, little information exists about how these pesticides might affect the gaseous products of the denitrification process, i.e. the N₂O-to-N₂ ratio.

Denitrification is a respiratory microbial process by which oxides of nitrogen serve as electron acceptors for respiratory electron transport in anaerobic conditions. As a result nitrate (NO₃⁻) is reduced to nitrite (NO₂⁻) and then gaseous products mainly N₂O and dinitrogen (N₂) (Knowles 1982; Simek *et al.* 2000). Denitrification has been studied intensively as it is an important contributor to the emission of N₂O. N₂O is a strong greenhouse gas with a global warming potential that is 300 times more powerful than carbon dioxide (for time horizon of 100 years) (IPCC 2001).

De novo synthesis of the reductases involved in denitrification process is inhibited by chloramphenicol while acetylene (C₂H₂) inhibits the activity of nitrous oxide reductase (Smith and Tiedje 1979). As such, the N₂O-to-N₂ ratio of the denitrification process can be determined when C₂H₂ is used while chloramphenicol at low

concentrations allows to study the denitrification capacity of the soil upon sampling (Knowles 1982; Pell *et al.* 1996). The objective of this study was to determine the effect of the pesticide methylparathion on dynamics of the denitrification process in a Chinampa soil using acetylene and chloramphenicol as selective inhibitors.

Methods

Soil samples were collected in the Chinampa of Xochimilco in San Gregorio Atlapulco (Xochimilco, Mexico City, Mexico).

As part of a study into the effects of methylparathion on soil processes one Chinampa soil was sampled. The sampling site of 4500 m² was divided in three equal plots. Soil was sampled by augering the 0-15 cm layer of the three plots with a stony soil auger diameter 7 cm (Eijkelkamp, NL). The soil of each plot was pooled and sieved (5 mm). Sub-samples of soil were amended with or without methylparathion and incubated aerobically at 22±2°C for 28 days. At the onset of the experiment and every seven days, soil was amended with or without acetylene (C₂H₂) or chloramphenicol and incubated anaerobically for 48 h while dynamics of NO₃⁻, NO₂⁻ and N₂O were monitored, after 0, 6, 24 and 48 h, three sub-samples of each treatment (*n* = 4), plot (*n* = 3) and amended with or without pesticide (*n* = 2) were selected at random for assays of NO₃⁻, NO₂⁻ and N₂O. The headspace of each flask was sampled and analysed for N₂O. The concentrations of N₂O were corrected for gas dissolved in the water (Moraghan and Buresh 1977). After measurement of N₂O, samples were analysed for NO₂⁻ and NO₃⁻ as described earlier. Data for NO₂⁻ were corrected for the formation of NO₂⁻ through the degradation of chloramphenicol (Dendooven *et al.* 1994).

Emission of N₂O was regressed on elapsed time using a linear regression model which was forced to pass through the origin but allowed different slopes (production rates) for each treatment. This approach is supported by theoretical considerations that no N₂O was produced at time zero and the atmosphere in the flask contained no N₂O as the headspace was flushed with N. Production of NO₂⁻ and NO₃⁻ was regressed on elapsed time using a linear model that was not forced to pass through the origin and allowed different slopes (production rates) for each treatment.

Results

The concentration of NO₃⁻ decreased over time (Table 1). In the SMP soil, day of sampling or treatment had no significant effect on the decrease in concentration of NO₃⁻ (Table 1). In the CMP soil, the decrease in the concentration of NO₃⁻ was slowest in soil conditioned aerobically for 7 days and the fastest for soil incubated for 28 days, while treatment had no significant effect on it. Methylparathion had no significant effect on the concentration of NO₃⁻ in soil (Table 2).

No clear pattern emerged in the concentrations of NO₂⁻ during the different anaerobic incubation. In the SMP soil, the concentration of NO₂⁻ (mean of all treatments) was significantly larger at day 0 and 7 than at day 14 and 28. In CMP soil, the mean concentration of NO₂⁻ was significantly larger at day 7 than at day 14 and 28, but significantly lower than at the onset of the experiment (*p* < 0.05) (Table 1). In the SMP soil, the mean concentration of NO₂⁻ was largest when amended with C₂H₂ plus chloramphenicol and lowest in the unamended SMP soil. In CMP soil, the concentration of NO₂⁻ increased significantly when amended with chloramphenicol compared to soil not amended with chloramphenicol. The concentration of NO₂⁻ was significantly larger in the CMP soil than in the SMP soil (*p* < 0.05) (Table 2).

Table 1. Effect of time of aerobic conditioning at 25 °C, i.e. 0, 7, 14 and 28 days, and treatment, i.e. control, acetylene, chloramphenicol, acetylene+chloramphenicol, on mean concentrations of NO₂⁻ (mg N/kg soil) and NO₃⁻ and N₂O production rates (mg N/kg soil/h) in soil amended or not with methylparathion incubated anaerobically at 25 °C for 48 h.

Time of conditioning	NO ₂ ⁻ concentration (mg N/kg soil)		Concentration of NO ₃ ⁻		Emission of N ₂ O	
	SP ^a	CP ^b	SP	CP	SP	CP
Day 0	.3 A ^c	8.8 A	-0.39 A	-0.61 BC	0.014 A	0.032 B
Day 7	.1 A	.4 B	-0.27 A	-0.11 A	0.011 B	0.043 A
Day 14	.4 B	.7 C	-0.26 A	-0.26 AB	0.009 B	0.022 C
Day 28	.9 B	.2 C	-0.42 A	-0.83 C	0.010 B	0.016 C
LSD ^d	.8	.0	SEE ^e 0.19	0.19	0.001	0.003
Treatment						
Control	.1 C	.5 B	-0.23 A	-0.33 A	0.007 B	0.020 B
Acetylene (C ₂ H ₂)	.2 B	.4 B	-0.25 A	-0.50 A	0.014 A	0.038 A
Chloramphenicol	.8 BC	1.9 A	-0.39 A	-0.53 A	0.008 B	0.018 B
Chloramphenicol+(C ₂ H ₂)	.6 A	2.3 A	-0.42 A	-0.59 A	0.014 A	0.034 A
LSD (P<0.05)	.8	.0	SEE 0.19	0.21	0.001	0.003

^a SP: without methylparathion,

^b CP: with methylparathion,

^c Values with the same capital letter are not significantly different within the column at $p < 0.05$,

^d LSD: Least significant difference ($p < 0.05$),

^e SEE: Standard error of the estimates ($p < 0.05$).

The emission of N₂O increased over time and resembled a zero order kinetic. However in the CMP soil amended with C₂H₂, the emission of N₂O increased after 24 h. In the SMP soil, the N₂O emission rate was significantly larger at the onset of the experiment than when conditioned aerobically for 7, 14 or 28 days (Table 1). In CMP soil, the highest N₂O emission rate was found when conditioned aerobically for 7 days and lowest when incubated aerobically for 14 or 28 days. The emission of N₂O was significantly larger in the CMP soil than in the SMP soil ($p < 0.05$) (Table 2).

The production of N₂ increased over time and resembled a zero order kinetic. However, in CMP soil conditioned for 7 days, the emission of N₂ increased after 24 h. The emission of N₂ was significantly larger in the CMP soil than in the SMP soil ($p < 0.05$) (Table 2).

Table 2. Effect of methylparathion on mean concentrations of NO₂⁻ (mg N/kg soil) and NO₃⁻, N₂O and N₂ production rates (mg N/kg soil/h) of soil conditioned for 0, 7, 14 or 28 days and left untreated or treated with acetylene, chloramphenicol, acetylene+chloramphenicol, and incubated anaerobically at 25 °C for 48 h.

	NO ₂ ⁻ concentration (mg N/kg soil)	NO ₃ ⁻ production rate	N ₂ O emission	N ₂ emission	N ₂ emission + chloramphenicol
- Methylparathion	9.9 B ^a	-0.19 A	0.011 B	0.009 B	0.005 B
+ Methylparathion	13.2 A	-0.54 A	0.028 A	0.015 A	0.012 A
LSD ^b (P<0.05)	1.6	SEE ^c 0.22	0.001	0.002	0.001

^a Values with the same capital letter are not significantly different within the column at $p < 0.05$,

^b LSD: Least significant difference ($p < 0.05$),

^c SEE: Standard error of the estimates ($p < 0.05$).

Conclusions

Pesticides inhibit or stimulate the denitrification process. Methylparathion when added to a Chinampa soil stimulated the denitrification process. It increased the concentration of NO₂⁻ and removal of NO₃⁻ from soil, and increased the emission of N₂O and N₂. Additionally, methylparathion did not affect the N₂O-to-N₂ ratio.

References

- CICOPLAFEST (2008) Control del Proceso y Uso de Plaguicidas, Fertilizantes y Sustancias Tóxicas. (<http://www.sagarpa.gob.mx/cicoplafest/>).
- Dendooven L, Splatt P, Anderson JM (1994) The use of chloramphenicol in the study of the denitrification process: some side-effects. *Soil Biology & Biochemistry* **26**, 925-927.
- INECOL (2002) Final report In: Programa rector de restauración ecológica área natural protegida zona sujeta a conservación ecológica 'Ejidos de Xochimilco y San Gregorio Atlapulco' (<http://ramsar.conanp.gob.mx/documentos/fichas/50.pdf>).
- IPCC (2001) Climate Change (2001) The Scientific Basis. Contribution of working group I to the third assessment report of the intergovernmental panel on climate change. (Eds JT Houghton, Y Ding, DJ Griggs, M Noguer, PJ van der Linden, X Dai, K Maskell, C.A Johnson). 881 pp. (Cambridge University Press, Cambridge).
- IPCC (2007a) Climate Change 2007: The Physical Science Basis. Contribution of working group I to the fourth assessment report of the intergovernmental panel on climate change. (Eds S Solomon, D Qin, M Manning, Z Chen, M Marquis, K.B Averyt, M Tignor, H.L Miller) 996 pp. (Cambridge University Press).
- IPCC (2007b) Climate Change 2007: Mitigation of Climate Change. Contribution of working group III to the fourth assessment report of the intergovernmental panel on climate change. (Eds B Metz, OR Davidson, PR Bosch, R Dave, LA Meyer.) pp. 499-532. (Cambridge University Press)
- Johnsen K, Jacobse SS, Torsvik V, Sorensen J (2001) Pesticide effects on bacterial diversity in agricultural soils. *Biology and Fertility of Soils* **33**, 454-459.
- Knowles R (1982) *Microbiological Reviews* **46**, 43-70.
- Locke MA, Zablotowicz RM (2004) Pesticides in Soil – Benefits and limitations to soil health. In 'Managing Soil Quality: Challenges in Modern Agriculture' (Eds. P Schjonning, S Elmholt, BT Christensen) pp. 239-260. (CABI Publishing)
- Rojas RT (1983) La agricultura chinampera. Compilación histórica. Dirección de difusión cultural. Cuadernos universitarios. *Agronomía* **7**, 181-211.
- Simek M, Cooper JE, Picek T, Santruckova H (2000) Denitrification in arable soils in relation to their physico-chemical properties and fertilization practice. *Soil Biology & Biochemistry* **32**, 101-110.
- Spokas K, Wang D (2003) Stimulation of nitrous oxide production resulted from soil fumigation with chloropicrin. *Atmospheric Environment* **37**, 3501-3507.
- Spokas K, Wang D, Venterea R (2005) Impact of soil fumigation with chloropicrin and methyl isothiocyanate on greenhouse gases. *Soil Biology & Biochemistry* **37**, 475-485.
- Spokas K, Wang D, Venterea R, Sadowsky M (2006) Mechanisms of N₂O production following chloropicrin fumigation. *Applied Soil Ecology* **31**, 101-109.
- Zhang R, Jiang J, Gu JD, Li S (2006) Long term effect of methylparathion contamination on soil microbial community diversity estimated by 16S rRNA gene cloning. *Ecotoxicology* **15**, 523-530.